

RESEARCH ARTICLE



Detection of *Ureaplasma Urealyticum* in Clinical Samples from Infertile Women by Polymerase Chain Reaction

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ABSTRACT

Genital Ureaplasma urealyticum infection is considered to be a sexually transmitted infection. The bacterium has been found to be involved in PID, chorioamnionitis, urethritis, respiratory distress syndrome and pneumonia in newborn, abortion and infertility. U. urealyticum infections not only jeopardize fertility but also pose a risk for infertility treatment and resulting pregnancies. The purpose of this study was to determine the prevalence of U. urealyticum in specimens from infertile women by polymerase chain reaction (PCR). 377 endocervical swab samples were taken from infertile women. Mycoplasma genus and U. urealyticum were detected by means of the polymerase chain reaction with specific primers. Mycoplasma genus DNA was detected in specimens from 141(37.4%) of 377 patients. Of these 141 patients 85(22.5% of total specimens) were PCR positive with urease gene primers for U. urealyticum. The isolation rate of U. urealyticum in young women (<30 age) was higher than the others. Because of the potential adverse effects of U. urealyticum on the success rate of highly specialized infertility treatment, and its causal roles in several maternal complications of pregnancy and in neonatal morbidity and mortality, the rapid detection of U. urealyticum by PCR in infertile women could be important and necessary. The increased sensitivity and shorter time requirement of PCR support its further development for the diagnosis of U. urealyticum infections.

Keywords: Ureaplasma urealyticum, Mycoplasma, Infertility, PCR

Mycoplasmas are the smallest cell free-life microorganisms. They can be isolated as commensals or pathogens from plants, insects, animals and humans. Some of them are considered normal flora of the respiratory or genitourinary tract [1]. U. urealyticum can be found in the cervix or vagina of 40-80% of sexually mature, asymptomatic women [2-6]. The presence of U.urealyticum in a large proportion of healthy women complicates the assessment of the pathogenic roles of this organism, but several studies have indicated that genital colonization of the U.urealyticum can be associated with an increased risk of developing certain pathogenic conditions and pregnancy abnormalities, e.g., pelvic inflammatory disease, premature rupture of membranes, chorioamnionitis, and preterm labor and birth. In addition, it may be acquired by neonates either in utero or by vertical transmission at birth and can cause pneumonia, pulmonary hypertension, chronic lung disease, and meningitis of the newborn [7-12].

During the past decade, evidence has accumulated of causative role of *U. urealyticum* in human infertility. U.urealyticum was detected at a higher frequency in infertile women [13-15]. Colonization of the upper female genital tract with *U. urealyticum* was found to be associated with adverse pregnancy out comes [16]. In human in vitro fertilization systems, the presence of *U. urealyticum* in either semen or the female genital tract resulted in a decrease in pregnancy rate per embryo transfer [17].

Diagnosis of mycoplasm is usually made by serological determination or in vitro isolation of the organism. However, serological procedures are often hampered by interspecies cross-reaction, while cultivation is time-consuming and hard to for some fastidious mycoplasmas. Use of mycoplasma species-specific DNA probes made it possible to discriminate between different species, this method proved to be rapid and specific [18].In this study we determined the prevalence of U.

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Table 1. Detection of Mycoplasma genus and U. urealyticum by PCR

PCR result	Number of specimens		Total number of
	PCR positive	PCR negative	specimens
Mycoplasma genus	141(37.4%)	236	377
U. urealyticum	85(22.5%)	292	377

urealyticum in endocervical samples from infertile women by polymerase chain reaction (PCR).

METHODS

Clinical specimens

Endocervical swab samples were taken from a total of three hundred and seventy seven infertile women, ranging in age from 17 to 45 years. Swab samples were placed immediately in sterilized container with 2ml of PBS (0.1M Nacl, 2.5mM Kcl, 10mM Na₂HPo₄, 1.5 mM KH₂Po₄, PH 7.4) for subsequent PCR.

DNA extraction from specimens

DNA was extracted from standard strain *U.urealyticum* serotype VIII (type strain) and clinical samples as described previously by Cadieux et al [19]. Briefly, 1ml of the sample was centrifuged at 12000 ×g for 10 min. The pellet was washed in PBS and resuspended in 30µl of distilled water. After boiling for 10 min, an aliquot of 7µl was used directly in PCR experiments.

PCRs

Mycoplasmas have been detected previously with genus-specific primers, followed by amplification of positive samples with species-specific primers for U. urealyticum. Mycoplasma-specific 16S rRNA fragments amplified by use of the published [20] Mycoplasma genus specific primers GSO (5-GGGAGCAAACAGGAT TAG ATACCCT-3) and MGSO (5 -TGCACCATCTGTCACTCTGTTAACCTC-3) The PCR assay was performed in 50ul of reaction mixture containing 10ul of 10× PCR buffer, 2.5 mM MgCl₂, 200 µM dNTP, 1.25 units of Taq polymerase, 20 pmol of each primer and 7µl of sample DNA. The reaction mixtures were placed in thermal cycler (Eppendorf, USA). The thermal profile involved an initial denaturation step at 94°C for 3 minutes followed by 30 cycles of denaturation at 94°C for 1 minute, primer annealing at 64°C for 1 minute, and primer elongation at 72°C for 1 minute. The cycling was followed by a final extension step at 72°C for 10 minutes. The primers published by Blanchard et al [21] were used for identification of U. urealyticum: primers U5 (5-CAATCTGCTCGTGAA GT ATTAC-3) and U4 (5-ACGACGT CCATAAGCAACT-3). The samples were placed in the same thermal cycler and heated to 94°C for 3 minutes. The cycling profile consisted of 30 cycles of 94°C(denaturation), 52°C(annealing), and

Table 2. Detection of U. urealyticum from patients according to age

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Age (years)	17-27	28-37	38-47	Total
Number of specimen	181	164	32	377
PCR posi- tive	45(24.8%)	34(20.7%)	6(18.7%)	85(22.5%)

 $72^{\circ}C(\text{elongation})$ for 1 minute. The cycling was followed by a final extension step at $72^{\circ}C$ for 10 minute. Aliquots of amplified samples (10μ l) were analyzed by electrophoresis on a 1% agarose gel stained with ethidium bromide.

Statistical analysis

Chi-square (X²) test was used for the generation of p < 0.05 values.

RESULTS

The DNA from each sample was subjected to two PCRs. The first PCR with primers GSO and GMSO, were based on genus-specific 16S rRNA gene sequences. The genus-specific primers reacted with all mycoplasmal species investigated as well as with members of the genera *Ureaplasma, Spiroplasma, and Acholeplasma* [20]. Of the 377 patients studied, 141(37.4%) were positive with genus-specific primers (Table 1). Of these 141 patients, 85 (22.54% of total samples) were PCR positive with species-specific primers for *U. urealyticum* (second PCR).

A photograph of electrophoresis based on bromidestained agarose gel for PCR-amplified products from the *Mycoplasma* and *Ureaplasma* strains is presented in figure 1. DNA from the 16S rRNA sequences that is amplified by the PCR primers used in this study shows at 270bp [20]. A 429bp fragment of the urease gene was amplified for identification of *U. urealyticum*. They have been shown previously to be highly specific for *U. urealyticum* and under optimal conditions, to allow detection of <10CFU of each serotype the organism [21].

The age of the patients from who were PCR positive varied from 17-45 with a mean age of 31 years. Distribution of the genital *U. urealyticum* in accordance to patient's age is presented in table 2. NO significant difference was found between the age of patients whose sample was PCR positive (positive group) and that of the other patients (negative group). There was also no difference regarding the duration of infertility, vaginal discharge, cervicitis, and abortion, between the positive and the negative group.

DISCUSSION

In this study, 22.54% of 377 infertile women were colonized with *U. urealyticum* as detected by PCR. Other studies using PCR for detecting *U. urealyticum* in endocervical specimens have reported prevalence rate as high as 40 to 80%.



Fig 1. Electrophoretic analysis of PCR products for Mycoplasma genus and U.urealyticum from endocervical samples; Lane 1: 100bp size marker, lane 2: standard strain of Mycoplasma genus (270bp), lane 3&4: patients positive samples for Mycoplasma genus, lane 5: standard strain of *U.urealyticum* (429bp), lane 6&7: patients' positive samples for *U.urealyticum*, lane 8: negative control (distilled water).

Since *U. urealyticum* has been found significantly associated with low socioeconomic background, such as poverty, number of sexual partners, and use of contraceptive drug [2-6], it is not surprising that the rate of *U. urealyticum* was lower in our study. Although differences were not statistically significant, but the isolation rate of *U. urealyticum* was higher in women under 30 years of age, that is consistent with that previously described by others [2,3,5,22].

At present, the main method of detecting *U. urealyticum* is by culture, but the cultivation of *U. urealyticum* is laborious, time consuming, and requires specific expertise. PCR is revolutionizing the diagnosis of many infectious diseases, particularly those caused by organisms that are difficult to cultivate. PCR is a more sensitive and reliable means of detecting *U. urealyticum* in the endocervical specimens; its results can be available within a day, compared with 2-5 days for culture [23].

Although the precise role of *U. urealyticum* in human infertility has not firmly established, there is strong support in the literature for causal role in the cases of women and men infertility [12-17,24-29]. In addition, *U. urealyticum* has been implicated in several maternal complications of pregnancy and in neonatal morbidity and mortality. It may plays roles in endometraitis, chorioamnionitis, premature rupture of membranes, prematurity, low-birth-weight infants, postpartum fever, and it is important causes of pneumonia and meningitis in very low-birth-weight infants. Therefore the specificity, the exquisite sensitivity, and the rapidity of PCR make this technique most valuable in the diagnosis of genital *U. urealyticum* infections.

References

 Razin S, Yogev D, and Naot Y. Molecular biology and pathogenicity of mycoplasmas. Microbiol Mol Biol Rev 1998; 62:1094-156.

- Domingues D, Tavira LT, Duarte A, et al. Genital mycopasmas in women attending a family planning clinic in Guine-Bissau and their susceptibility to antimicrobial agents. Acta Tropica 2003; 86: 19-24.
- Clegg A, Passey M, Michael A. High rates of genital mycoplasma infection in the highlands of Papua New Guine determined both by culture and by a commercial detection kit. J Clin Microbiol 1997; 35: 197-200.
- Fourmaux S, Bebear C. Urogenital infection linked to chlamydia and mycoplasmas. Prog Urol 1997; 7: 132-6.
- Gratlard F, Soleihac B. Epidemiologic and molecular investigations of genital mycoplasmas from women and neonates at delivery. Pediatr Infect Dis J 1995; 14: 853-8.
- keane FE, Thomas BJ, Gilroy CB, et al. The association of Mycoplasma hominis, Ureaplasma urealyticum, and Mycoplasma genitalium with bacterial vaginosis: bservations on hetrosexual women and their male partners. Int J STD AIDS 2000; 11: 356-60.
- Roberston JA, Honore LH, Stemke GW, et al. The association of parvo biotype of Ureaplasma urealyticum with cases of reproductive failure. J O M Lett 1992; 2: 63-5.
- Yoon BH, Romero R, Kim M ,et al. Clinical implications of detection of Ureaplasma urealyticum in the amniotic cavity with the Polymerase chain reaction. Am J Obstet Gynecol 2000; 183: 1130-7.
- Yoon BH, Romero R, Park JS, et al. Fetal exposure to an intraamniotic inflammation and the development of cerebral palsy at the age of three years. Am J Obstet Gynecol 2000; 182: 675-681.
- Grether JK, and Nelson KB. Maternal infection and cerebral palsy in infants of normal brith weight. JAMA,1997, 287:207-211
- Cedillo-Ramirez I, Gil C, Zago I, et al. Association of Mycoplasma hominis and Ureaplasma urealyticum with some indicator of non-specific vaginitis. Rev.Latinoam.2000, 42:1-6
- Donders GG, Van Bulk B, Caudrom I, et al. Relationship of bacterial vaginosis and mycoplasmas to the risk of spontaneous abortion. Am.J.Obstet.Gynecol. 2000,183: 431-437
- Gnarpe H, and Friberrgy M. Mycoplasma and human reproductive failure. Am.J.Obstet.Gynecol.1972,114, 727-731
- Charpe H, and Friberrgy M. T- mycoplasmas as a possible cause for reproductive failure. Nature. 1973,242: 120-121
- Taylor-Robison D. Evaluation of the rate of Ureaplasma urealyticum in infertility. Pediatr.Infect.Dis. 1989, 5(suppl): 262-265
- Andrew WW,Goldenberg RI, Hauth JC. Preterm labor: emerging role of genital tract infections. Infect. Agents Dis 1995; 4: 196-211.
- 17. Montagut JM, Lepretre S, Degoy J, et al. Ureaplasma urealyticum in semen and IVF. Hum Reprod 1991; 61: 727-729.
- Kuppeveld FJM, Logt JTM, Angulo AF et al. Genus and species-specific identification of Mycoplasmas by 16S rRNA amplification. Appl Env Microbiol 1992; 52: 2606-2615.
- Cadieux N, Lebel P, and Brousseau R. Use of a triplex polymerase chain reaction for the detection and differentiation of Mycoplasma pneumoniae and Mycoplasma genitalium in the presence of human DNA. J Gen Microbiol 1993; 139: 2431-2437.
- Kuppeveld FJ, Johansson KE, Galama JM etal. Detection of mycoplasma contamination in cell cultures by a Mycoplasma group-specific PCR. Appl Env Microbiol 1994; 4: 149-152

- Blanchard A, Henstehel J, Duffy L, et al. Detection of Ureaplasma urealyticum by polymerase chain reaction in the urogenital tract of adults, in amniotic fluid, and in the respiratory tract of newborns. Clin Infect Dis 1993; 17(suppl 1): 48-53.
- 22. leon X. The presence of gentital mycoplasmas in women of reproductive age. Rev Med Panama 1993; 18: 238-241.
- Comparison of PCR with culture for detection of Ureaplasma urealyticum in clinical samples from patients with urogenital infections. J Clin Microbiol 1994; 32: 2232-34.
- Nunez-Calonge R, Caballero P, Redondo C, Baquero F, Martinez-Ferrer M, and Meseguer MA. Ureaplasma urealyticum reduces motility and induces membrane alterations in human spermatozoa. Hum Reprod 1998;13: 257-261.
- 25. Witkin SS, Kligman J, Grifo JA, and Rosenwaks Z. Ureaplasma urealyticum and Mycoplasma hominis detected by the polymerase chain reaction in the cervices of women undergoing in vitro fertilization :prevalence and consequences. J.Assist .Reprod Genet.1995; 12: 610-614
- 26. Malka R, Kahane I and Bartov B. In vitro and In vivo impairment of human and Ram sperm nuclear chromatin integrity by

sexually transmitted Ureaplasma urealyticum infection. Biol.reprod 2000; 63: 1041-1048

- 27. Fenkci V, Yilmazer M, Aktepe OC. Have Ureaplasma urealyticum and Mycoplasma hominis infection any significant effect on women fertility ? Infez.Med 2002; 10: 220-223
- Shalika S, Dugan K, Smith RD, and Padilla SL. The effect of positive semen bacterial and Ureaplasma cultures on in vitro fertilization success. Hum Reprod1996;11:2789-2792
- 29. Kanokas N, Mantzavinos T, Boufidou F, Koumentakou I, and Creatsas G. Ureaplasma urealyticum in semen: is there any effect on in vitro fertilization? Fretil steril 1999;71:523-527.

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