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ORIGINAL ARTICLE

Comparative Evaluation of in Vitro Effects of Praziquantel (PZQ) on the Enzyme Activities of the Excretory-Secretory Products (ESP) of Fasciola hepatica and F. gigantica Parasites

ALI FARAHNAK, TAGHI GOLMOHAMDI, and MOHAMAD REZA ESHRAGHIAN

For author affiliations, see end of text.

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ABSTRACT

Excretory-secretory products (ESP) play an important role in the host biochemical defense by means of activities of detoxifying and antioxidant enzymes. The aim of this study was to evaluate praziquantel (PZQ) effects by detection of glutathione S- transferase (GST) and superoxide dismutase (SOD) enzymes activities in ESP samples of Fasciola spp. Fasciola gigantica and Fasciola hepatica adult parasites were collected and cultured within buffer media for 4 h at 37°C. Treated (with 50, 100 and 150 ug PZQ) and control ESP samples for each species were collected and centrifuged. The supernatants were stored at -20°C. SOD and GST enzymes activities of ESP samples were estimated photometrically. To determine the statistically-significant difference between ESP of treated and control samples, t-test was conducted. ESP protein bands were detected by gel electrophoresis (SDS-PAGE). SOD enzyme activities level in treated F.hepatica and F. gigantica ESP samples were determined as 0.26, 0.29, 0.76 and 1.73, 1.60, 1.43 U/ml respectively. SOD activities level in control samples were detected 1.31 and 1.35 U/ml. GST activities level in treated F.hepatica and F. gigantica ESP samples were calculated 9.4, 3.1, 23.96 and 435.4, 333.3, 720.9 U/ml. GST activities levels in control samples were detected 43.8 and 134.4 U/ml respectively. Statistical analysis revealed the significant decrease of GST and SOD enzyme activities in treated ESP samples of Fasciola hepatica, and significant increase of GST enzyme activity in treated ESP samples of F. gigantica in comparison to the control samples (p < 0.05). There was also no difference between SDS-PAGE results of treated and control samples. Based on the results of present work, PZQ has decreasing effect on the ESP enzymatic activities of Fasciola hepatica and increasing effect on F. gigantica enzyme activities. In other words, F. hepatica has less capability to protect against xenobiotics and free radicals than F.gigatica does.

Keywords: Fasciola gigantica, F.hepatica; Excretory-Secretory Products, Praziquantel, Superoxide dismutase, Glutathione S- transferase

Millions of people are infected with fascioliasis in the world. The disease is caused by trematodes belonging to Fasciola (*F. hepatica* and *F. gigantica*) parasites and treated with triclabendazole [1]. At the present time, praziquantel is not using for treatment of fascioliasis; however, this anthelmintic drug is very effective against other parasitic diseases [2-3].

Various studies have been recorded on the characterization of fasciola parasite ESP [4-5]. Glutathione S-transferase (GST) and superoxide dismutase (SOD) enzymes are common in the ESP of fasciola parasite [6]. The antioxidant enzyme, SOD, is capable of inhibiting the oxidation of other molecules [7]. In the other hand, GST enzymes is detoxifying

Table 1. SOD enzyme activities value of praziquantel (PZQ)-treated excretory- secretory product (ESP) samples of *Fasciola hepatica*; *F. gigantica* and control samples (mean of 10 ESP samples for each)

ESP Samples	SOD* activities of <i>F. gigantic</i> (U/ml)**	SOD* activities of <i>F. hepatica</i> (U/ml)**
Control ESP samples	1.35	1.31
ESP contains 50 mg PZQ	1.73	0.26
ESP contains 100 mg PZQ	1.60	0.29
ESP contains150 mg PZQ	1.43	0.76

^{*}Superoxide dismutase Enzyme

Table 2. GST enzyme activities value of praziquantel (PZQ) treated excretory- secretory product (ESP) samples of *Fasciola hepatica*; *F. gigantica* and control samples (mean of 10 ESP samples for each)

ESP Samples	GST* activities of F. gigantic (U/ml)**	GST* activities of <i>F. hepatica</i> (U/ml)**
Control ESP samples	134.4	43.8
ESP contains 50 μg PZQ	435.4	9.4
ESP contains 100 μg PZQ	333.3	3.1
ESP contains 150 μg PZQ	720.9	23.9

^{*}Glutathione S- transferase enzyme

endogenous com pounds as well as breaking down the xenobiotics [8].

Although there are many studies on the effects of triclabendazole on the fasciola products, however, there are no reports on PZQ effects on fasciola products [9-11]. In this research, in order to evaluate PZQ effects on GST and SOD enzyme activities, the PZQ-treated ESP of *Fasciola hepatica* and *F.gigantica* were compared with control samples in culture.

MATERIALS AND METHODS

Collection of parasite ESP samples

The adult parasites of *Fasciola gigantica* and *Fasciola hepatica* were collected from local abattoir (Tehran abattoir, Tehran, Iran) and identified, based on morphologic and morphometric characters. Collected alive Fasciola parasites were washed for a minimum of three times in PBS, pH 7.4, to remove host material. Meanwhile, to prepare PZQ dilutions (50, 10, 150 µg/ml), purchased PZQ tablets (Tolid Darou Dami Iran Co.) was homogenized within ethanol as a solvent. The parasites were cultured within the buffer media (treated or control) for 4 h at 37°C (2 parasites in 2 ml of culture media for each sample). The total of 80 ESP samples of *Fasciola gigantica* and *Fasciola hepatica* (30 treated, 10 for each PZQ dilution, and 10 control ESP samples for each species) were collected and centrifuged at

10000 g for 30 min and supernatants were stored at -20°C [12].

SOD enzyme activity assay of ESP samples

SOD activity of ESP samples were determined using RANSOD Kit (Randox Labs, crumlin, UK). The absorbances of samples were measured 30 s after the addition of xanthine oxidase as start reagent at 505 nm on a spectrophotometer (Cecil Instruments Ltd) and 3 min after reaction, in 37°C A standard curve was plotted using the standard provided in the Kit and the SOD total activity value in U/ml for each ESP sample was read from this curve using the excel software. One unit enzyme activity is the amount of SOD that inhibits the rate of formazan dye formation by 50% [13].

GST enzyme activity assay of ESP samples

For activity assay of GST enzyme in ESP samples, reagent cocktail including, potassium phosphate buffer, reduced glutathione (GSH) and 1-chloro-2, 4-dinitrobenzene (CDNB) substrates were prepared in the cuvette. In each cuvette, from the mentioned mixture, 200 µl sample solution was removed and then the same volume of PZQ-treated *F. hepatica* and *F. gigantica* ESP or control samples was added into cuvette and mixed well. Finally, the cuvette was placed into the barrel of spectrophotometer and absorbances recorded for 5 minutes at 340 nm. Total GST enzyme activity as U/ml of all samples were calculated. One unit GST activity

^{**}One unit enzyme activity is the amount of SOD that inhibits the rate of formazan dye formation by 50%

^{**}One unit will conjugate 1.0 µmole of 1-chloro-2, 4-dinitrobenzene with reduced glutathione per minute at pH 6.5 at 25°C

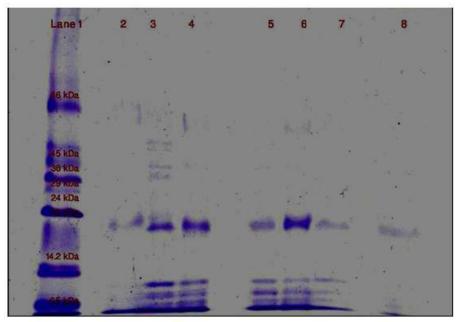


Fig 1. SDS- PAGE analysis of PZQ-treated and control ESP samples from F. hepatica and F. gigantica. Each lane represents an ESP sample; the lane 1 corresponds to the protein marker. The lanes 2, 3 and 4 correspond to treated F. hepatica and the lanes 5, 6 and 7 correspond to treated F. gigantica ESP samples by PZQ dilutions of 50, 100, 150 µg/ml. The lane 8 indicates control ESP sample.

conjugate 1.0 µmole of 1-chloro-2, dinitrobenzene with reduced glutathione per minute at pH 6.5 at 25°C [14].

SDS-PAGE analysis of ESP samples

Sodium dodecyl sulfate Polyacrylamide electrophoresis and coomassie blue staining were used to separate the components of ESP samples protein. Samples were added to each wells of gel, 7.5 %, and were run for 6 hours at 15 mA and finally the gels were stained by comassie blue staining. Molecular weights of sample proteins were detected using the protein marker within the gel [12].

Statistical analysis of ESP samples

To determine the statistical difference between GST and SOD values of enzyme activities in PZQ-treated and control ESP samples, two-sample independent ttests was used [15].

RESULTS

The results of SOD and GST enzyme activities assay are presented in Tables 1 and 2. In F. hepatica, two-sample t-test results reveal that there is significant decrease of SOD and GST activities of treated ESP samples in comparison to the control ESP samples (p <0.05). In F. gigantic, t-test results show that there is significant increase of GST activities of treated ESP samples in comparison to the control ESP samples (p <0.05). However, there is no significant difference between SOD activities in treated F. gigantica and control ESP samples (p > 0.05).

Protein bands of ESP samples were detected using SDS electrophoresis (Fig 1). There is no difference

between SDS-PAGE results of treated and control ESP samples.

DISCUSSION

GST and SOD enzyme activities in ES of Fasciola hepatica and F. gigantica were previously demonstrated by authors [16-17]. Pentoxifylline as adjuvant therapy with praziquantel produced reduction in glutathione-Stransferase (GST) and superoxide dismutase (SOD) in experimental Schistosomiasis mansoni [18]. In this study, the significant decrease of GST enzyme activity in PZQ-treated F.hepatica ESP may be due to less ability of this species to detoxifies the endogenous compounds as well as breakdown of xenobiotics in comparison to F.gigantica. This phenomenon increase parasite attack in the liver tissue of man and animals by the endogenous compounds. On the other hand, the significant increase of GST enzyme activity in treated F. gigantica ESP may be due to more ability of this species to detoxify endogenous compounds as well as breakdown of xenobiotics in comparison with the F.hepatica. At the present research, the significant decrease of SOD enzyme activity of treated Fasciola hepatica ESP increase molecular oxidation and consequently assists the parasite destruction in the liver tissue. Although there is no significant difference for SOD activities of treated F. gigantica and control ESP samples, higher activity of this enzyme causes more protection of fasciola gigantic than F. hepatica dose against free radical.

In conclusion, the significant decrease of GST and SOD enzyme activities in PZQ-treated Fasciola hepatica ESP samples indicate this species has less capability to protect against xenobiotics and free radicals than *F. gigatica* does.

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CURRENT AUTHOR ADDRESSES

- Ali Farahnak, Department of Parasitology and Mycology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran. E-mail: farahnak@tums.ac.ir (Corresponding Author)
- Taghi Golmohamdi, Department of Biochemistry, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran.
- Mohamad Reza Eshraghian, Department of Epidemiology and Biostatistics, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran.