Antidiarrhoeal and Antiulcer Activities of *Mammea Africana*

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**ABSTRACT**

The stem bark of *Mammea africana* is used traditionally for the treatment of stomach pains. The present study was designed to evaluate the effects of ethanolic stem bark extract of *Mammea africana* on experimentally induced diarrhoea and ulcer. The extract (30 – 90mg/kg) was administered orally in rats 30 minutes to one hour before the induction of diarrhoeal and ulcer using different experimental models. Castor oil (2ml) was used to induce diarrhoea, while indomethacin (60mg/kg) and ethanol (2.5ml/kg) were used to induce ulcers in rats. Frequency of defecation, inhibition of intestinal propulsion and fluid accumulation were measured in the case of diarrhoea and ulcer index were calculated to examine ulcer preventive ratio of the extract. The extract (30 – 90mg/kg) significantly inhibited castor oil induced diarrhoea, small intestine transit time and castor oil induced fluid accumulation as well as indomethacin and ethanol induced ulcer models. The effect of the extract in these models was uncomparable to the various standard drugs used. These findings justify the use of this stem bark traditionally in the treatment of stomach pains and disorders.

**Keywords:** *M. African, Stomach pains, Diarrhoea*

*Mammea africana* sabine (Guttiferae) (syn. *Ochrocarpus africana* Oliv.) is a large forest tree of 50 to 100 feet high with bark often yellow with pale scales and resinous yellow sap [1]. The plant is widely distributed in tropical Africa. The stem bark of the plant is used traditionally by the Ibibios of Niger Delta region of Nigeria in the treatment of malaria related fever, diabetes, and microbial infections. The stem bark is also used traditionally to treat stomach pains, rheumatic pains, scabies, cough and hypertension[2,3]. The chloroformic and ether stem bark extract are reported to possess cytotoxic activity on cell culture [4]. Ouahouo et al. [5], reported cytotoxic coumarins with antimicrobial activity against *Staphylococcus aureus* from the plant stem bark. The stem bark has been reported to possess antimalarial [6], cardioprotective [7], antidiabetic and hypoglycaemic [8], vasorelaxant [9], antihypertensive [10] effects. The stem bark has been reported to contain 5,7-dihydroxy-8-(12-methyl-butryl) -4- N - Pentyl coumarins [11,12,13], Mesuxanthone B [11]. Alkaloids have been reported to be absent in the entire plant parts [14]. Although reports of scientific studies on *Mammea africana* have been widely published, there is no information regarding the antidiarrhoeal and antiulcer activities of the stem bark extract of the plant in rats eventhough it is used traditionally in the treatment of gastrointestinal tract disorders.

The present study, therefore, was to establish if the stem bark of *M. africana* has any antiulcer and antidiarrhoeal activities to confirm its ethnomedical uses in the treatment of stomach disorders.

**MATERIALS AND METHODS**

*Plant materials*

Fresh stem bark of *M. africana* were collected in November, 2007 at Anwa forest in Uruan, Akwa Ibom State, Nigeria. The plant was identified and authenticated by Dr. Margaret Bassey, a taxonomist in the Department of Botany, University of Uyo, Uyo, Nigeria. Herbarium specimen was deposited at Faculty of Pharmacy Herbarium (voucher no. FPHUU. 381). The fresh stem bark (2 kg) of the plant were was on laboratory table for 2 weeks and reduced to powder. The powder 100g was macerated in 95% ethanol (300ml) for 72 hours [6]. The liquid filtrate obtained was concentrated in vacuo at 40°C. The yield was 2.08% w/w. The extract was stored in a refrigerator at 4°C until used for experiment reported in this study.
Animals

Albino wistar rats (105 – 165g) of either sex used for the experiments were obtained from the University of Uyo animal house. They were housed in standardized environmental conditions (22 ± 2.5 °C, relative humidity 80 – 85%, 12h light/12h dark cycle) and maintained on standard animal pellets and water ad libitum. Permission and approval for animal studies were obtained from the College of Health Sciences Animal Ethics committee, University of Uyo.

Castor oil induced diarrhoea

Diarrhoea was induced in adult albino male rats using a modified method developed by Sunil et al., [15, 16]. Animals were fasted for 24h but allowed free access to water. They were randomized into five groups of six rats each. Group 1(control) received 10% Tween 80 (5ml/kg) orally, Group 2- 4 were treated with M. africana extract (30, 60 and 90mg/kg, p.o. respectively); Group 5 was treated with atropine (0.1mg/kg, i.p) and Group 6 treated with extract, 60mg/kg, ten minutes later atropine 0.1mg/kg, i.p). After 1h, each rat received 2ml of castor oil (p.o) and was then observed for consistency of faecal matter and frequency of defecation for 3h.

Small intestinal propulsion

The effects of the extract on intestinal propulsion in unanaesthetized adult albino male rats were tested using charcoal method of Nwafor and Okwuasaba, [17]. Adult albino male rats were fasted for 24h but allowed free access to water only and were further randomized into six groups of six rats each. Group 1(control) received 10% Tween 80 (5ml/kg) by orogastric gavage; group 2-4 were treated with M. africana extract (30, 60 or 90mg/kg, p.o. respectively); Group 5 received atropine (0.1mg/kg, i.p) and Group 6 treated with extract, 60mg/kg, ten minutes later atropine 0.1mg/kg, i.p). After 1h, each rat received 2ml of castor oil (p.o) and was then observed for consistency of faecal matter and frequency of defecation for 3h.

Castor oil induced fluid accumulation.

Fluid accumulation was induced in adult albino male rats according to the method of DiCarlo et al., [18]. Animals were deprived of food for 24h but allowed free access to water. They were randomized into 6 groups of six rats each. Group 1(control) received castor oil (2ml/rat), Group 2 – 4 were administered with M. africana (30, 60 or 90 mg/kg, p.o), Group 5 received atropine (0.1mg/kg, i.p), Group 6 received atropine (0.1mg/kg,i.p), 10min later, M. africana extract (60mg/kg, p.o.). After 1h, each rat received 2ml castor oil (p.o.), 30 minutes later, the rats were killed by cervical dislocation and exsanguinated, the small intestine was ligated at both pyloric sphincter and at the ileocaecal junctions. The entire small intestine was dissected out, its content was expelled into a graduated measuring cylinder and the volume of the contents recorded.

Indomethacin induced ulcer

Male adult albino rats were used for the experiment. They were randomized into six groups of six rats each. Food was withdrawn 24 hours and water 2h before the commencement of experiment [19]. Group 1(control) received only indomethacin (Sigma, 60mg/kg p.o. dissolved in 5% Na2CO3); Groups 2- 4 were pretreated with Mammea africana extract (30, 60 and 90 mg/kg p.o. respectively)Group 5 received cimetidine (100mg/kg p.o. dissolved in 50% Tween 80), while Group 6 received cimetidine (100mg/kg. p.o) and 10 minutes later, extract (60mg/kg. p.o) was given. One hour later, groups 2 - 6 were administered with indomethacin. Four hour after indomethacin administration, animals were killed by cervical dislocation. The stomachs were removed and opened along the greater curvature. The tissues were fixed with 10% formaldehyde in saline. The tissues were fixed with 10% formaldehyde in saline. Macroscopic examination was carried out with a hand lens and the presence of ulcer lesion was scored [20]. Ulcer index (UI) and preventive ratio (PR) of each of the groups pretreated with extract was calculated using standard methods [21, 22].

Ethanol induced gastric ulceration

The procedure was similar to that used in indomethacin induced ulceration. Adult albino male rats were randomly assigned into six groups of six rats each. Food was withdrawn 24 hours and water 2h before the commencement of experiment [19]. Group 1(control) received only ethanol (2.5 ml/kg p.o),Groups 2- 4 were pretreated with Mammea africana extract (30,60, and 90mg/kg p.o. respectively); Group 5 received propranolol (40mg/kg p.o. dissolved in distilled water), while Group 6 received propranolol (40mg/kg. p.o dissolved in distilled water) and 10 minutes later, extract (60mg/kg,p.o) was given. One hour later, groups 2 - 6 were administered with ethanol. Four hour after ethanol administration, animals were killed by cervical dislocation. The stomachs were removed and opened along the greater curvature. The tissues were fixed with 10% formaldehyde in saline. Macroscopic examination was carried out with a hand lens and the presence of ulcer lesion was scored [22].
Table 1. Effect of *Mammea africana* extract on castor oil induced diarrhea in rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (mg/kg)</th>
<th>Mean faecal matter</th>
<th>%inhibition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (10% Tween 80)</td>
<td>-</td>
<td>8.76±0.91</td>
<td>-</td>
</tr>
<tr>
<td><em>M. africana</em> extract</td>
<td>30</td>
<td>4.66±0.75*</td>
<td>46.80</td>
</tr>
<tr>
<td></td>
<td>60</td>
<td>5.00±0.77**</td>
<td>42.92</td>
</tr>
<tr>
<td></td>
<td>90</td>
<td>5.00±1.09**</td>
<td>42.92</td>
</tr>
<tr>
<td>Atropine</td>
<td>0.1</td>
<td>0.66±0.42***</td>
<td>92.46</td>
</tr>
<tr>
<td><em>M. africana</em> + atropine</td>
<td>60+0.1</td>
<td>1.33±0.55***</td>
<td>84.81</td>
</tr>
</tbody>
</table>

Data were expressed as mean ± SEM. significant at *p*<0.01, **p<0.001 when compared to control n = 6.

Table 2. Effect of *Mammea africana* extract on small intestinal propulsion in rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (mg/kg)</th>
<th>Intestinal transit%</th>
<th>%inhibition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (10% Tween80)</td>
<td>-</td>
<td>67.2±0.21</td>
<td>-</td>
</tr>
<tr>
<td><em>M. africana</em> extract</td>
<td>30</td>
<td>64.6±0.90</td>
<td>3.86</td>
</tr>
<tr>
<td></td>
<td>60</td>
<td>63.6±0.24</td>
<td>5.36</td>
</tr>
<tr>
<td></td>
<td>90</td>
<td>56.3±0.84**</td>
<td>16.22</td>
</tr>
<tr>
<td>Atropine</td>
<td>0.1</td>
<td>41.8±0.91**</td>
<td>37.79</td>
</tr>
<tr>
<td><em>M. africana</em> + atropine</td>
<td>60+0.1</td>
<td>60.66±2.60*</td>
<td>9.73</td>
</tr>
</tbody>
</table>

Data were expressed as mean ± SEM. significant at *p*<0.001, **p<0.001, when compared to control n = 6.

Table 3. Effect of *Mammea africana* extract on castor oil induced fluid accumulation in rats

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (mg/kg)</th>
<th>Mean volume of intestinal fluid (ml)</th>
<th>% reduction</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (10% Tween80)</td>
<td>-</td>
<td>2.40±0.19</td>
<td>-</td>
</tr>
<tr>
<td><em>M. africana</em> extract</td>
<td>30</td>
<td>1.80±0.26</td>
<td>25.00</td>
</tr>
<tr>
<td></td>
<td>60</td>
<td>1.90±0.14</td>
<td>20.83</td>
</tr>
<tr>
<td></td>
<td>90</td>
<td>1.40±0.17*</td>
<td>41.66</td>
</tr>
<tr>
<td>Atropine</td>
<td>0.1</td>
<td>0.80±0.10**</td>
<td>66.66</td>
</tr>
<tr>
<td><em>M. africana</em> + atropine</td>
<td>60+0.1</td>
<td>0.80±0.14**</td>
<td>66.66</td>
</tr>
</tbody>
</table>

Data were expressed as mean ± SEM. significant at *p*<0.01, **p<0.001, when compared to control n = 6.

**Statistical analysis**

Data obtained from this study were statistically analysed with one-way ANOVA, followed by Tukey–Kramer multiple comparison post test. Values of *p*<0.01 were considered significant.

**RESULTS**

**Castor oil-induced diarrhoea**

*Mammea africana* extract (30 – 90mg/kg) reduced the castor oil-induced diarrhoea significantly (*p*<0.01). However, the effect of the median dose (60mg/kg) was similar to that of the highest dose (90mg/kg). The effect of the extract was lower than that of the standard drug, atropine, used in the study. The effect of the extract was enhanced in the presence of atropine, a muscarinic antagonist (Table 1).

**Castor oil induced intestinal propulsion**

Table 1 shows the effect of *M. africana* stem bark extract on intestinal propulsion of rats. The extract inhibited the intestinal propulsion in a dose-dependent manner. However, the effect was only significant (*p*<0.001) at the highest dose of the extract (90mg/kg) The effect of the extract was enhanced in the presence of atropine, a muscarinic antagonist (Table 2).

**Castor oil induced intestinal fluid accumulation**

The stem bark extract (30 – 90mg/kg) demonstrated a significant reduction in intestinal fluid accumulation due to castor oil administration relative to control. The reduction, though dose dependent was significant (*p*<0.01) only at the highest dose of the extract (90mg/kg) when compared to control but lower than that of the standard drug, atropine, used in the study. The effect of the extract was enhanced in the presence of atropine, a muscarinic antagonist (Table 3).

**Indomethacin induced gastric ulceration**

The extract pretreatment on indomethacin – induced gastric ulceration exerted a statistically significant and dose dependent decrease (*p*<0.01) in ulcer indices relative to control. The effect of the extract was comparable to that of the standard drug, cimetidine (Table 4).

**Ethanol induced gastric ulceration**

The stem bark extract pretreatment significantly (*p*<0.001) reduced the ulcer indices of ethanol induced ulceration relative to control. The effect of the extract was more than that of the standard drug, propranolol (40mg/kg) (Table 5).

**DISCUSSION**

In this study, ethanolic stem bark extract of *M. africana* exhibited significant antidiarrhoeal and antiulcer activities in the models tested. Castor oil induces diarrhea due to active ingredient, ricinoleic acid, which is liberated as a result of action of lipases on castor oil. This stimulates peristaltic activity in the small intestine, leading to changes in the electrolyte...
permeability of the intestinal mucosa. It also stimulates the release of endogenous prostaglandins [23, 24]. Castor oil elicits secretory and motility diarrhoea [25]. Inhibitors of prostaglandin synthesis are known to delay diarrhoea induced with castor oil [15]. The observations suggest that the antidiarrhoeal effect of the extract may be due to inhibition of prostaglandin synthesis. Also since the effect was enhanced in the presence of atropine, the extract may have acted also through antimuscarinic activity to reduce castor oil induced diarrhea. The extract also exhibited a significant inhibition of the small intestine propulsive movement, but the effect was not comparable to that of the standard drug, atropine, used in the study. More so, when the extract was given with atropine, an anticholinergic, enhanced activity was observed pointing to a possible involvement of anticholinergic activity. Conversely, the extract may be inhibiting the small intestinal movement through some other mechanism such as antagonism of α2 adrenoceptor stimulation. This extract also inhibited significantly (p<0.001) castor oil induced intestinal fluid accumulation (enteropooling). This effect was not comparable to that of the standard drug, atropine, which also exerts antisecretory activity. The effect of the extract, however, was enhanced in the presence of atropine, supporting earlier suggestion that the extract may be acting through an anticholinergic mechanism. Antidiarrhoeal and antidisentric properties of medicinal plants were found to be due to the presence of tannins, alkaloids, saponins, flavonoids, steroids and or terpenoids [26]. Coumarins, flavonoids and xanthones have been reported to be present in the stembark extract of this plant [11,12,13,27]. Coumarins, flavonoids and xanthones have been reported to cause relaxation of vascular smooth muscles [28,29], though acting through different mechanisms. These constituents which have been reported to be present in the extract of the *M. africana* maybe responsible for the *in vivo* antidiarrhoeal activity of *M. africana* stembark extract.  

**Table 4. Effect of *M. africana* extract on indomethacin induced ulcer**

<table>
<thead>
<tr>
<th>TREATMENT</th>
<th>DOSE (mg/kg)</th>
<th>ULCER INDICES</th>
<th>PREVENTIVE RATIO</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (indomethacin)</td>
<td>60</td>
<td>12.3±0.14</td>
<td>-</td>
</tr>
<tr>
<td><em>M. africana</em> extract p.o.</td>
<td>30</td>
<td>7.8±0.33*</td>
<td>39.0</td>
</tr>
<tr>
<td></td>
<td>60</td>
<td>7.50±2.21</td>
<td>36.3</td>
</tr>
<tr>
<td></td>
<td>90</td>
<td>4.00±0.49b</td>
<td>67.5</td>
</tr>
<tr>
<td>Cimetidine</td>
<td>100</td>
<td>4.16±1.51a</td>
<td>66.2</td>
</tr>
<tr>
<td>Cimetidine + <em>M. africana</em></td>
<td>100+60</td>
<td>3.83±0.47b</td>
<td>68.7</td>
</tr>
</tbody>
</table>

Data were expressed as mean ± SEM. significant at p < 0.001, 0.01, 0.05 when compared to control n = 6.

**Table 5. Effect of *Mammea africana* extract on ethanol induced ulcer**

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (mg/kg)</th>
<th>Ulcer indices</th>
<th>Preventive ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (ethanol)</td>
<td>-</td>
<td>5.76±0.47</td>
<td>-</td>
</tr>
<tr>
<td><em>M. africana</em> extract p.o</td>
<td>30</td>
<td>2.33±0.43*</td>
<td>59.5</td>
</tr>
<tr>
<td></td>
<td>60</td>
<td>2.04±0.02*</td>
<td>64.6</td>
</tr>
<tr>
<td></td>
<td>90</td>
<td>1.32±0.12*</td>
<td>77.1</td>
</tr>
<tr>
<td>Propranolol</td>
<td>40</td>
<td>3.00±0.01*</td>
<td>47.9</td>
</tr>
<tr>
<td>Propranolol + <em>M. africana</em></td>
<td>40 + 60</td>
<td>0.50±0.22*</td>
<td>91.3</td>
</tr>
</tbody>
</table>

Data were expressed as mean ± SEM. significant at *p< 0.001 when compared to control n = 6.

*Mammea africana* stembark extract was also evaluated for antiulcer activity using indomethacin and ethanol – induced ulcer models. Indomethacin, a known ulcerogen especially on an empty stomach [30] causes ulcer mostly on the glandular (mucosal) part of the stomach [20, 31], by inhibiting prostaglandin synthetase through the cycloxygenase pathway [32]. Prostaglandins function to protect the stomach from injury by stimulating the secretion of bicarbonate and mucus, maintaining mucosal blood flow and regulating mucosal turnover and repair [33, 34]. Suppression of prostaglandin synthesis by indomethacin results in increased susceptibility stomach to mucosal injury and gastro duodenal ulceration. The extract was observed to significantly reduce mucosal damage in the indomethacin – induced ulcer model, suggesting the possible extract mobilization and involvement of prostaglandin in the anti ulcer effect of the extract. Administration of ethanol has been reported to cause disturbances in gastric secretion, damage to the mucosa, alterations in the permeability, gastric mucus depletion and free radical production [35]. This is attributed to the release of superoxide anion and hydroperoxy free radicals during metabolism of ethanol as oxygen derived free radicals have been found to be involved in the mechanism of acute and chronic ulceration in the gastric mucosa [36]. It was observed in this study that the extract significantly reduced ethanol- induced ulcer. This may be due to cytoprotective effect of the extract via antioxidant effects. Ethanol is also reported to cause gastric mucosal damage by stimulating the formation of leukotriene C4 (LTC4) [37]. The gastroprotective effect of the extract may in part be due to the suppression, by the extract of lipoxygenase activity [20]. Okokon et al. [6], reported that the stembark extract contains flavonoids, terpenes, saponins, alkaloids and cardiac glycosides among others. Flavonoids such as querectin have been reported to prevent gastric mucosal lesions in various experimental models [38, 39] by increasing the
amount of neutral glycoproteins [38]. Flavonoids have been reported to protect the gastric mucosa from damage by increasing the mucosal prostaglandin content and by inhibiting histamine secretion from mast cells by inhibition of histidine decarboxylase. Free radical scavenging ability of flavonoids has been reported to protect the gastrointestinal tract from ulcerative and erosion lesion [39]. Saponins, especially triterpenes type have been implicated in antiulcer activity mediated by formation of protective mucus on the gastric mucosa and also protect the mucosa from acid effects by selectively inhibiting PGF2α [40, 41].

In conclusion, the results of this study have shown that the stem bark extract of M. africana possess antiulcer and antiulcerative activity and this confirms the ethnomedical usage of the stem bark extract in the treatment of stomach pains.

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REFERENCES


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