

Free Radical Scavenging Activity of Some Plants Available in Malaysia

ZAINUL AMIRUDDIN ZAKARIA

For author affiliations, see end of text.

Received February 21, 2007; Accepted May 29, 2007

This paper is available online at <http://ijpt.iums.ac.ir>

ABSTRACT

The present study was carried out to determine the free radical scavenging properties of some plants found in Malaysia such as, *Muntingia calabura*; *Bauhinia purpurea*; *Dicranopteris linearis*; *Melastoma malabathricum*; *Corchorus capsularis*. The air-dried leaves of each plant (20 g) were soaked in distilled water (1:20; w/v) for 72 h at room temperature. The collected supernatants were tested for the free radical scavenging activity against the DPPH and superoxide anion radical scavenging assays. All extracts were found to show remarkable antioxidant activity in both assays with the percentage of inhibition (%) yielded 94–99% and 83–100%, respectively. Phytochemicals screening of all plants demonstrated the presence of flavonoids, saponins, triterpenes and steroids, but not alkaloids. Tannins was detected only in the leaves of *M. calabura*, *D. linearis*, *M. malabathricum*. The ability to scavenge free radicals indicates these plants could be used as a new source of antioxidant agents, and the activity seen could be attributed to the synergistic effect of various bioactive compounds present in these extracts, particularly of the flavonoids type. Further study has been designed in our laboratory to isolate and to identify the bioactive compounds responsible for the observed antioxidant activity.

Keywords: *Muntingia calabura*; *Bauhinia purpurea*; *Dicranopteris linearis*; *Melastoma malabathricum*; *Corchorus capsularis*; Aqueous extract; Antioxidant activity

Various forms of activated oxygen, generally known reactive oxygen species (ROS), have been implicated in many diseases, i.e. cancer, diabetes, atherosclerosis and heart disease [1]. ROS, which can be classified into free radicals (i.e. superoxide ions ($O_2^{\cdot-}$) and hydroxyl radicals ($OH\cdot$)) and non-free-radicals (hydrogen peroxide (H_2O_2)) [2,3], are produced from endogenous sources within the living organisms via various mechanisms (i.e. normal aerobic respiration, stimulated poly-morpho-nuclear leukocytes and macrophages, and peroxisomes) or from exogenous sources (tobacco smoke, ionizing radiation, certain pollutants, organic solvents, and pesticides) [4-6]. Free radicals, generated *in vivo* due to the various biochemical reactions occurring in the living tissues, are chemical species that have tendency to rob electrons from other molecules in the immediate surrounding in order to replace their own losses. This process will lead in damage of crucial bio-molecules including those present in cell membranes, mitochondria, DNA, etc. and thus predisposing various pathophysiological states if not effectively scavenged. Although tissue injury leads to the generation of ROS, the ROS

can also cause tissue injury when present in high concentration within the tissues/cells [7].

It is well known that all aerobic organisms possess antioxidant mechanisms to protect against the oxidative damages, and various types of enzymes responsible for the removal or repair of the damaged molecules [8]. Even so, these natural mechanisms can be ineffective sometimes and therefore, dietary intake of antioxidants is essential [9,10]. The fact that synthetic antioxidants (i.e. butylated hydroxytoluene and butylated hydroxyanisole), commonly used in processed foods, possessed some side effects have limited their use as antioxidant agents [11,12]. Studies have demonstrated a converse relationship between the consumption of antioxidant-rich plants/vegetables and the incidence of human diseases [13]. Thus, research to find the new sources of natural antioxidants is important. Also, in recent years, attentions have been directed towards the possible therapeutic potential of antioxidants in controlling degenerative diseases associated with marked oxidative damage. Various plants have been shown to possess significant antioxidant property [14-16] and differ-

Table 1. The Free Radical Scavenging Activity of Aqueous Extracts of Several Plants Found in Malaysia

Aqueous Sample	DPPH Radical Scavenging (%)	Superoxide Scavenging (%)
<i>M. calabura</i>	94.80 ± 1.14	83.70 ± 2.05
<i>D. linearis</i>	96.60 ± 0.60	89.40 ± 0.80
<i>M. malabathricum</i>	98.30 ± 0.75	96.80 ± 1.25
<i>B. purpurea</i>	94.90 ± 1.05	100.00 ± 0.00
<i>C. capsularis</i>	97.50 ± 1.06	94.90 ± 0.50

ent classes of phytochemicals have been demonstrated to be responsible for the plants' antioxidant activity [17-19]. The aim of the present study was to determine the antioxidant activity of the leaves aqueous extracts of five plants available in Malaysia, namely *Muntingia calabura*, *Dicranopteris linearis*, *Melastoma malabathricum*, *Bauhinia purpurea* and *Corchorus capsularis*.

MATERIALS AND METHODS

Plant Material

The leaves of *M. calabura*, *D. linearis*, *M. malabathricum* and *B. purpurea* were collected from its natural habitat in Shah Alam, Selangor, Malaysia whereas the leaves of *C. capsularis* were collected from its natural habitat in Alor Setar, Kedah, Malaysia, between June and September 2005. They have been identified by Mr. Shamsul Khamis, a botanist from the Institute of Bioscience, Universiti Putra Malaysia, Malaysia and voucher specimens have been deposited at the Herbarium of the Laboratory of Natural Products, Institute of Bioscience, UPM, Serdang, Selangor, Malaysia.

Preparation of Aqueous Extract of Plants

The leaves of all plants were washed and rinsed with tap water and then oven-dried for 72 hours at 40°C. The dried leaves were then ground into small particles, weighed (40 g) and soaked for 72 h in distilled water (dH₂O) (1:20; w/v). The supernatant of each plant was collected and filtered using Whatman No. 1 filter paper and then subjected to the free radical scavenging assays.

DPPH Radical Scavenging Activity

Assay for DPPH free radical scavenging potential is based on the scavenging activity of stable DPPH free radicals [15]. Reaction mixtures containing test samples

dissolved in methanol and 200 µM DPPH (Sigma) in ethanolic solution in a 96-well microtiter plate were incubated at 37 °C for 30 min. After the reaction, absorbance was then measured at 520 nm, and percent inhibition was calculated.

Superoxide Anion Radical Scavenging Activity

The superoxide anion radical scavenging activity was performed using the method of Okamura et al. [16] with some modification. This assay is based on the removal rate of xanthine/xanthine oxidase-generated superoxide by measuring the reduction of nitro blue tetrazolium (NBT). The sample solution (0.1 mg/mL) in 5% DMSO was added to 1 mL of a mixture of 0.1 mM xanthine and 0.2 mM NBT (Sigma) in 50 mM potassium phosphate buffer (pH 7.5) containing 0.05 mM EDTA. Xanthine oxidase (0.1 mL) (Sigma, USA; 0.8 unit/mL) diluted in 50 mM phosphate buffer (pH 7.5) was added, and the resulting mixture was incubated at 37°C for 20 min. Addition of 2 mL of 2.5 N HCl to the mixtures terminated the reaction, followed by increase of coloration of NBT, which was measured at 540 nm. The percent of removal rate by sample was calculated relative to the control.

Phytochemical Screening of the Leaves of *M. calabura*, *D. linearis*, *M. malabathricum*, *B. purpurea* and *C. capsularis*

The phytochemical screening of the selected plants' leaves was carried out according to the standard screening tests and conventional protocols as described by Ikhiri et al. [20].

HPLC Profiling of the Leaves Aqueous Extracts of *M. calabura*, *D. linearis*, *M. malabathricum*, *B. purpurea* and *C. capsularis*

HPLC profiling of the plants' leaves aqueous extracts were carried out in the Laboratory of Phytomedicine, Forest Research Institute of Malaysia, Kepong,

Table 2. The Phytochemical Screening of Several Plants Found in Malaysia

Constituents	<i>M. calabura</i>	<i>D. linearis</i>	<i>M. malabathricum</i>	<i>B. purpurea</i>	<i>C. capsularis</i>
Flavonoids	+++	+	++	+	++
Triterpenes	++	+	++	+	+
Tannins	+	+	++	-	-
Alkaloids	-	-	-	-	-
Saponins	+	+++	+	++	+
Steroids	+++	+++	+++	+++	++

For flavonoids, tannins, triterpenes and steroids - - - weak colour; ++ - mild colour; +++ - strong colour

For saponins - - - 1-2 cm froth; ++ - 2-3 cm froth; +++ - >3 cm froth

For alkaloids - - - negligible amount of precipitate; ++ - weak precipitate; +++ - strong precipitate

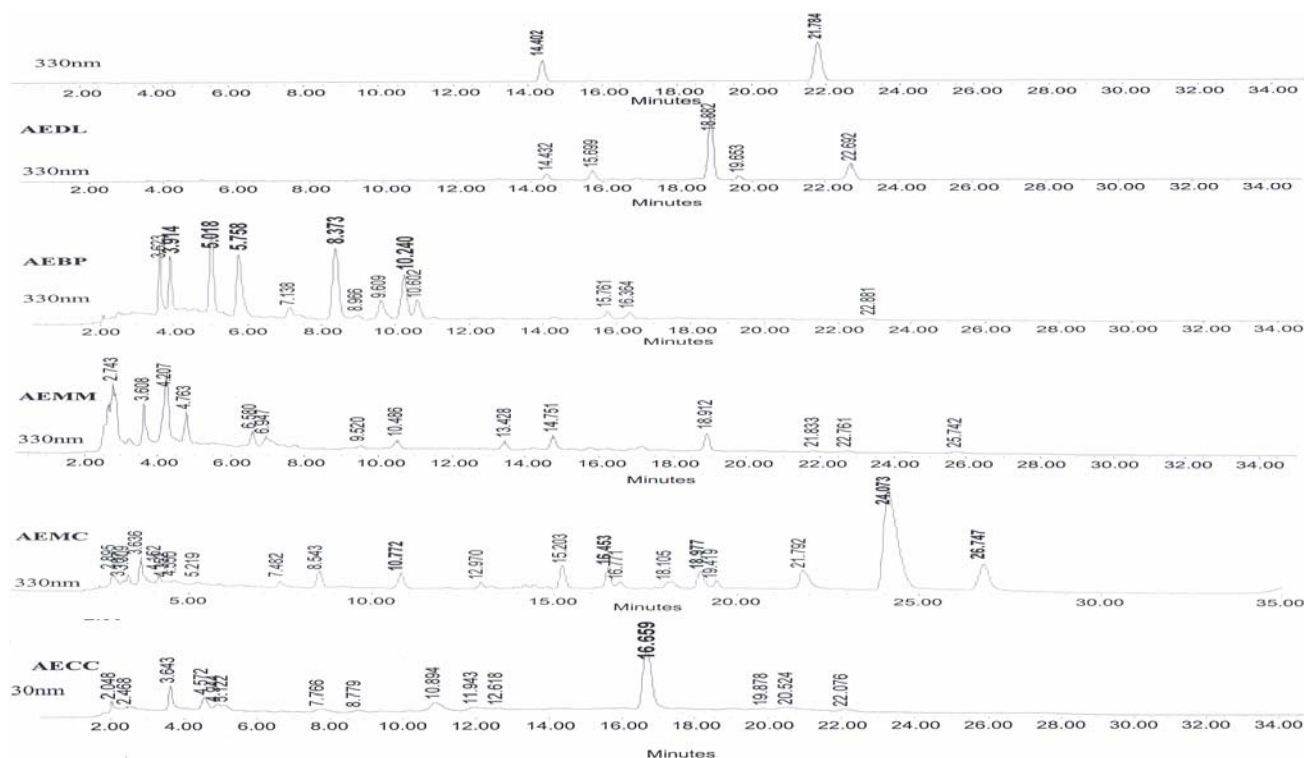


Fig 1. HPLC Profiles of the Selected Plants at 300 nm (AEDL – Aqueous extract of *D. linearis*; AEBP – Aqueous extract of *B. purpurea*; AEMM – Aqueous extract of *M. malabathricum*; AEMC; Aqueous extract of *M. calabura*; AECC – Aqueous extract of *C. capsularis*)

Malaysia. A Waters Delta 600 with 600 Controller and Waters 2996 Photodiode Array (Milford, MA, USA) equipped with an autosampler, online degasser and column heater was used for HPLC analysis. Data were analysed and processed using the installed Millenium 32 Software (Waters Product). The sample were separated at 27°C on a minibore Phenomenex Luna 5µm C₁₈ column with dimensions 250 x 4.60 mm using a one step linear gradient. The solvents were (A) 0.1% ortho-phosphoric acid and (B) acetonitrile. The elution system was as follows: 0-12 min, 0-15% B; 12-22 min, 15-25% B; 22-35 min, 25-15% B with a flow rate of 1000 µL/min. The HPLC was monitored at 330 nm and compared with standards flavonoids, namely rutin and fisetin.

RESULTS

The free radical scavenging effect of aqueous extracts of *M. calabura*, *D. linearis*, *M. malabathricum*, *B. purpurea* and *C. capsularis*, assessed using the DPPH radical scavenging and xanthine/xanthine oxidase superoxide assays, are shown in Table 1. Interestingly all extracts demonstrated remarkable scavenging effects with activity recorded above 80% in both tests.

The phytochemical screenings of the selected plants are demonstrated in Table 2. All plants demonstrated the presence of flavonoids, triterpenes, saponins and high content of steroids, but no traces of alkaloids. Interestingly, only three plants namely, *M. calabura*, *D. linearis* and *M. malabathricum*, showed the presence of tannins.

Fig 1 demonstrated the HPLC profiles of the aqueous extracts of selected plants. Comparison made at the wavelength of 330 nm against standard flavonoids, namely rutin (retention time (R_T) = 14.402 min) and catechin (R_T = 21.786 min), indicate that all extracts possessed R_T that is different from the standards, except for the AEDL (R_T = 14.432 min) and AEMC (R_T = 21.792 min). Although the retention time could not give confirmation on the actual compounds that might be present in the extracts, it can be used as a comparison against standards, which have been known to possess a specific retention time. Further study; however, need to be carried out in order to obtain the actual bioactive compounds responsible for the extracts free radical scavenging activities.

DISCUSSION

The present study demonstrated the radical scavenging property of several plants, namely *M. calabura*, *D. linearis*, *M. malabathricum*, *B. purpurea* and *C. capsularis*, found in Malaysia. The aqueous extracts of the selected plants' leaves, assayed against the DPPH radical scavenging and xanthine/xanthine oxidase superoxide assays, were found to exhibit remarkable radical scavenging activities with the percentage of inhibition recorded for the former and latter assays ranged between 94 – 99% and 83 – 100%, respectively.

It is generally known that the antioxidant activities of putative antioxidants involves various mechanisms, such as radical scavenging, decomposition of peroxides, binding of transition metal ion catalysts, prevention of chain initiation and prevention of continued hydrogen

abstraction [21]. Hence, the free radical scavenging capacity of an extract may serve as a significant indicator of its potential antioxidant activity. Increasing evidences have suggested that many age-related human diseases (i.e. cancer, inflammation and brain dysfunction) are the result of cellular damage caused by free radicals [22,23]. Antioxidants have been shown to play an important role in preventing such diseases. For example, several cancer chemopreventive agents exhibit antioxidant activity through their ability to scavenge oxygen radicals [24,25].

According to Winston [26], the leafy part of the vegetables/plants contains various types of bioactive compounds. In general, all of the plants used in this study demonstrated to contain flavonoids, saponins, triterpenes and steroids, but no alkaloids. In addition, tannins were only detected only in *M. calabura*, *D. linearis*, *M. malabathricum*. The interests in phenolic compounds, particularly flavonoids and tannins, have considerably increased in recent years because of their broad spectrum of chemical and diverse biological properties, which include the antioxidant effects [27] and radical scavenging properties [28]. Flavonoids have been associated with possible role in the prevention of several chronic diseases involving oxidative stress [29], as well as their protective effect against low-density lipoprotein (LDL) oxidation [30]. Thorough phytochemical investigations of the used plants and evaluation for their antioxidant activity by means of other models (i.e. various ex vivo and in vivo biochemical assays) have been planned in our laboratory to characterize the bioactive compounds with potential antioxidant activity.

All of the plants used in this study have been reported to possess anti-inflammatory and antinociceptive activities [31-35]. Studies have demonstrated the link between the anti-inflammatory and antinociceptive, and antioxidant activities of the plants. For example, nitric oxide (NO) is produced/released under the action of inflammatory stimuli (i.e. ROS) [36]. Inhibition of ROS leads to the reduction of NO production, which has been demonstrated to cause anti-inflammatory, antinociceptive and antioxidant activities [37,38]. The free radical scavenging property may be one of the mechanisms by which these plants' are effective in their ethnopharmacological uses against different ailments. In term of the flavonoids, much attention has been given to their antioxidant [39] and anti-inflammatory activities, in vitro and in vivo [40,41]. Flavonoids have been reported to inhibit cytokine (inflammatory stimuli) release from RAW264.7 cells [42] and may modulate the increasing number of cellular processes involving redox reaction, including the regulation of tyrosine phosphatase activity [43].

In conclusion, the present study provides preliminary evidence on the antioxidant property, partly *via* the free radical scavenging activity, of the leaves aqueous extract of several plants found in Malaysia, namely *M. calabura*, *D. linearis*, *M. malabathricum*, *B. purpurea* and *C. capsularis*. Flavonoids, triterpenes, saponins and

tannins are believed to act synergistically to produce the observed activity.

ACKNOWLEDGEMENTS

This study was supported by the research grant of Universiti Industri Selangor, Malaysia (Project Code Number: 03013; Project Vote Number: 3090103013). The authors would like to thank Universiti Putra Malaysia for the facilities.

REFERENCES

1. Hertog MGL, Feskens EJM, Hollman PCH, et al. Dietary anti-oxidant flavonoids and risk of coronary heart disease: The Zutphen elderly study. *The Lancet*. 1993; 342: 1007-1014.
2. Halliwell B. How to characterize an antioxidant: an update. *Biochem Soc Symp*. 1995; 61: 73-101.
3. Squadriato GI, Pelor WA. Oxidative chemistry of nitric oxide: The roles of superoxide, peroxynitrite, and carbon dioxide. *Free Rad Biol Med*. 1998; 25: 392-403.
4. Halliwell B, Gutteridge JM. *Free radicals in biology and medicine*. Clarendon Press Oxford 1989; 23-30.
5. Yildirim A, Oktay M, Bülaloğlu V. The antioxidant activity of the leaves of *Cydonia vulgaris*. *Turk J Med Sci*. 2001; 31: 23-27.
6. Davies KJA. Oxidative stress the paradox of aerobic life. *Biochem Symp*. 1994; 61: 1-34.
7. Robinson EE, Maxwell SRJ, Thorpe GHG. An investigation of the antioxidant activity of black tea using enhanced chemiluminescence. *Free Rad Res*. 1997; 26: 291-302.
8. Auroma OI. Free radicals, oxidative stress, and antioxidants in human health and disease *JAOCS* 1998; 75: 199-212.
9. Tanizawa H, Ohkawa Y, Takino Y, et al. Studies on natural antioxidants in citrus species I. Determination of antioxidative activities of citrus fruits. *Chem Pharm Bull*. 1992; 40: 1940-1942.
10. Sasaki S, Ohta T, Decker EA. Antioxidant activity of water soluble fractions of salmon spermary tissue. *J Agric Food Chem*. 1996; 44: 1682-1686.
11. Branien AL. Toxicology and biochemistry of butylated hydroxyanisole and butylated hydroxytoluene. *JAOCS* 1975; 52: 59-63.
12. Ito N, Fukushima S, Hassegawa A, et al. Carcinogenicity of butylated hydroxyanisole in F344 rats. *J Natl Can Inst*. 1983; 70: 343-347.
13. Rice-Evans CA, Sampson J, Bramley PM, et al. Why do we expect carotenoids to be antioxidants in vivo. *Free Rad Res*. 1997; 26: 381-398.
14. Kweon M-H, Hwang H-J, Sung H-C. Identification and Antioxidant Activity of Novel Chlorogenic Acid Derivatives from Bamboo (*Phyllostachys edulis*). *J Agric Food Chem*. 2001; 49: 4646-4655.
15. Chen Y, Wong M, Rosen R, et al. 2,2-Diphenyl-1-picrylhydrazyl radical scavenging active components from *Polygonum multiflorum* Thunb. *J Agric Food Chem*. 1999; 47: 2226-2228.
16. Okamura H, Mimura A, Yakou Y, et al. Antioxidant activity of tannins and flavonoids in *Eucalyptus rostrata*. *Phytochem*. 1993; 33: 557-561.
17. Nakatani N, Kayano S, Kikuzaki H, et al. Identification, quantitative determination, and antioxidative activities of chlorogenic acid isomers in prune (*Prunus domestica* L.). *J Agric Food Chem*. 2000; 48: 5512-5516.
18. Yan X, Suzuki M, Ohnishi-Kameyama M, et al. Extraction and identification of antioxidants in the rots Yacon (*Smallanthus sonchifolius*). *J Agric Food Chem*. 1999; 47: 4711-4713.

19. Baderschneider B, Winterhalter P. Isolation and characterization of novel benzoates, cinnamates, flavonoids, and lignans from Riesling wine and screening for antioxidant activity. *J Agric Food Chem*. 2001; 49: 2788-2798.
20. Ikhirri K, Boureima D, Dan-Kouloudo D. Chemical screening of medicinal plants used in the traditional pharmacopoeia of Niger. *Int J Pharmacog*. 1992; 30: 251-262.
21. Diplock AT, Charleux JL, Crozier-Willi G, et al. Functional food science and defence against reactive oxygen species. *Br J Nutr*. 1998; 80: S77-S112.
22. Perry G, Raina AKL, Nonomura A, et al. How important is oxidative damage? Lessons from Alzheimer's disease. *Free Rad Biol Med*. 2000; 28: 831-834.
23. Carr A, Frei B. The role of natural antioxidants in preserving the biological activity of endothelium-derived nitric oxide. *Free Rad Biol Med*. 2000; 28: 1806-1814.
24. Ito H, Miyake M, Nishitani E, et al. Antitumor promoting activity of polyphenols from *Cowania mexicana* and *Coleogyne ramosissima*. *Cancer Lett*. 1999; 143: 5-13.
25. Wei HC, Frenkel K. Relationship of oxidative events and DNA oxidation in SENCAE mice to in vivo promoting activity of phorbol ester-type tumor promoter. *Carcinogen*. 1993; 14: 1195-1201.
26. Winston JC. Health promoting properties of common herbs. *Am J Clin Nutr*. 1999; 70: 491S-499S.
27. Larson RA. The antioxidants of higher plants. *Phytochem*. 1988; 27: 969-978.
28. Agrawal PK. Carbon-13 NMR of Flavonoids. New York: Elsevier Press; 1989: 5-31.
29. Lee KW, Kim YJ, Lee HJ, et al. Cocoa has more phenolic phytochemicals and a higher antioxidant capacity than teas and red wine. *J Agric Food Chem*. 2003; 51: 7292-7295.
30. Silva FAM, Borges F, Guimaraes C, et al. Phenolic acids and derivatives: Studies on the relationship among structure, radical activity, and physicochemical parameters. *J Agric Food Chem*. 2000; 48: 2122-2126.
31. Zakaria ZA, Sulaiman MR, Mat Jais AM, et al. The involvement of L-arginine/nitric oxide/cyclic guanosine monophosphate pathway in *Muntingia calabura* aqueous extract antinociception in mice. *Fundam Clin Pharmacol*. 2006; 20: 365-372.
32. Zakaria ZA, Abdul Rahman NI, Loo YW, et al. Antinociceptive and anti-inflammatory activities of the chloroform extract of *Bauhinia purpurea* leaves in animal models. *Yakugaku Zasshi* 2007; Submitted.
33. Zakaria ZA, Abdul Ghani ZDF, Raden Mohd. Nor RNS, et al. Antinociceptive and anti-inflammatory activities of *Dicranopteris linearis* leaves chloroform extract in experimental animals. *Yakugaku Zasshi* 2006; 126(11): 1197-1203.
34. Zakaria ZA, Raden Mohd. Nor RNS, Hanan Kumar G, et al. Antinociceptive, anti-inflammatory and anti-pyretic properties of *Melastoma malabathricum* leaves aqueous extract in experimental animals. *Can J Physiol Pharmacol* 2007; Accepted.
35. Zakaria ZA, Sulaiman MR, Hanan Kumar G, et al. Antinociceptive and anti-inflammatory properties of *Corchorus capsularis* leaves chloroform extract in experimental animal models. *Yakugaku Zasshi* 2007; Accepted.
36. Olszanecki R, Gębska A, Kozłowski VI, et al. Flavonoids and nitric oxide synthase. *J Physiol Pharmacol*. 2002; 53: 571-584.
37. Middleton E Jr, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. *Pharmacol Rev*. 2000; 52: 673-751.
38. Robak J, Gryglewski RJ. Flavonoids are scavengers of superoxide anions. *Biochem Pharmacol*. 1988; 37: 837-841.
39. Edenharder R, Grunhage D. Free radical scavenging abilities of flavonoids as mechanism of protection against mutagenicity induced by tertbutyl hydroperoxide or cumene hydroperoxide in *Salmonella typhimurium* TA102. *Mutat Res*. 540: 1-18.
40. Calixto JB, Otuki MF, Santos ARS. Anti-inflammatory compounds of plant origin. Part I. Action on arachidonic acid pathway, nitric oxide and nuclear factor κ -B (NF κ B). *Planta Med*. 2003; 69: 973-983.
41. Calixto JB, Campos MM, Otuki MF, et al. Anti-inflammatory compounds of plant origin. Part II. Modulation of pro-inflammatory cytokines, chemokines and adhesion molecules. *Planta Med*. 2004; 70: 93-103.
42. Xagorari A, Roussos C, Papapetropoulos A. Inhibition of LPS-stimulated pathways in macrophages by the flavonoid luteolin. *Br J Pharmacol*. 2002; 136: 1058-1064.
43. Gamet-Payrastre L, Manenti S, Gratacap MP, Tulliez J, Chap H and Payrastre B. Flavonoids and the inhibition of PKC and PI 3-kinase. *Gen Pharmacol* 1999; 32: 279-286.

CURRENT AUTHOR ADDRESSES

Zainul Amiruddin Zakaria, Faculty of Pharmacy, Universiti Teknologi MARA, 40450 Shah Alam, Selangor, MALAYSIA. E-mail: shaza8174@yahoo.com